

# Evolutionary morphology of haptor anchors in monogenoids (Dactylogyridae) of marine catfish (Siluriformes: Ariidae) from the Atlantic coast of South America

## Research Article

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### Abstract

Exploring the phylogenetic signal of morphological traits using geometric morphometry represents a powerful approach to assess the relative weights of convergence and shared evolutionary history in shaping species' forms. We evaluated the phylogenetic signal in shape and size of ventral and dorsal haptor anchors of 10 species of monogenoids (*Hamatopeduncularia*, *Chauhanellus* and *Susanlimocotyle*) occurring in marine catfish (Siluriformes: Ariidae) from the Atlantic coast of South America. The phylogenetic relationships among these species were mapped onto the morphospaces of shape and size of dorsal and ventral anchors. Two different tests (squared change-parsimony and  $K_{mult}$ ) were applied to establish whether the spatial positions in the phylomorphospace were influenced by phylogenetic relationships. A significant phylogenetic signal was found between anchor form and parasite phylogeny. Allometric effects on anchor shape were non-significant. Phylogenetically distant species on the same host differed markedly in anchor morphology, suggesting little influence of host species on anchor form. A significantly higher level of shape variation among ventral anchors was also found, suggesting that the evolutionary forces shaping ventral anchor morphology may operate with differing intensities or exhibit distinct mechanisms compared to their dorsal counterparts. Our results suggest that phylogenetic relationships were a key driver of changes in shape (but not size) of anchors of monogenoids of South American ariids. However, it seems that the emergence of the digitiform haptor in *Hamatopeduncularia* and in some species of *Chauhanellus* played an important role in the reduction in anchor size and may cause secondary losses of anchors in other groups of monogenoids.

### Introduction

Monogenoidea Bychowsky, 1937 (Platyhelminthes) are primarily ectoparasites of fish (Whittington, 2005). These parasites have as a major taxonomic structure, the haptor, which plays a key role for attachment to the gills or body surface of the host (Bychowsky, 1957; Boeger and Vianna, 2006). This structure integrates sclerotized hard parts such as hooks, anchors and clamps or a combination of these elements. Importantly, many monogenoids exhibit high host specificity (Whittington *et al.*, 2000), which indicates a highly specific adaptation to parasitize particular fish species.

Monogenoidea has demonstrated to be an excellent model system for studying the evolutionary processes that have driven parasite diversification and diversity (Poulin, 2002). Different studies have used these parasites to investigate the processes leading to their diversification and speciation (Šimková *et al.*, 2002; Vanhove and Huyse, 2015), to elucidate the evolutionary association of hosts and parasites (Desdevises *et al.*, 2002; Šimková *et al.*, 2006; Šimková and Morand, 2008; Mendlová and Šimková, 2014; Vanhove *et al.*, 2015; Míguez-Lozano *et al.*, 2017; Rahmouni *et al.*, 2022; Seidlová *et al.*, 2022; Soares *et al.*, 2023a) and to explore the relationship between phenotype variation in attachment organs and factors such as phylogeny and host specificity (Vignon *et al.*, 2011; Sarabeev and Desdevises, 2014; Llopis-Belenguer *et al.*, 2015; Khang *et al.*, 2016; Rodríguez-González *et al.*, 2017).

Some of these studies have been based on linear measurements of haptor elements (e.g. Mladineo *et al.*, 2013; Kmentová *et al.*, 2020; Cruz-Laufer *et al.*, 2022). This approach can leverage published datasets, enabling the analysis of extensive data volumes (Cruz-Laufer *et al.*, 2022). However, a major limitation of linear measure-based morphometrics is the inherent fusion of size and shape information, leading to difficulties in disentangling these 2 aspects

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(Adams *et al.*, 2004). Geometric morphometrics addresses this issue effectively and additionally provides visualization tools, such as shape deformation grids, that facilitate the interpretation and communication of intricate shape changes (Adams *et al.*, 2013). Another advantage of geometric morphometrics is that shape analysis relies on homologous landmarks, ensuring that comparisons between individuals or taxa are based on corresponding anatomical points. While identification of landmarks is more time-consuming due to the relatively involved collection and processing of data, traditional morphometrics often rely on arbitrary or poorly defined measurements that make it difficult to assess homology (Adams *et al.*, 2004).

Geometric morphometrics have been successfully utilized to investigate the evolutionary processes that have shaped the diversification of a wide range of organisms, including plants (Liu *et al.*, 2015), fishes (Friedman *et al.*, 2019), mites (Kerschbaumer and Pflingstl, 2021) and parasites (Vignon *et al.*, 2011; Baillie *et al.*, 2019; Soares *et al.*, 2023a). In Monogonoidea, different studies, using geometric morphometric data of the haptor anchors, associated with molecular phylogenies have underscored the usefulness of this approach in tackling diverse evolutionary inquiries (Llopis-Belenguer *et al.*, 2015; Khang *et al.*, 2016; Rodríguez-González *et al.*, 2017; Rahmouni *et al.*, 2021; Soares *et al.*, 2023a).

In the present study, we integrate geometric morphometrics of haptor anchors and DNA sequences in a comparative phylogenetic context, in order to investigate the evolution of form (i.e. the combination of shape and size *sensu* Klingenberg, 2016) of 10 species from 3 monogenoid genera (*Hamatopeduncularia* Yamaguti, 1953, *Chauhanellus* Bychowsky & Nagibina, 1969 and *Susanlimocotyle* Soares, Domingues and Adriano, 2021) that parasitize Ariidae (Siluriformes) from South America. Haptor anchors were chosen for analysis because they are not subjected to large variation due to contraction or flattening on fixation (Vignon, 2011) and are crucial for effective attachment to the host. In fact, Šimková *et al.* (2002) indicate that the morphology of the haptor is, to a large degree, determined by adaptation to the host and to attachment to specific sites within their hosts, which has been demonstrated in, for instance, *Lamellodiscus* spp. (Poisot *et al.*, 2011). Thus, similarity in anchor morphology could result from homoplasy, indicating convergent evolution. However, shared evolutionary history can also play a major role in determining anchor shape, as shown in *Ligophorus* spp. (Rodríguez-González, 2017). So, anchor morphology is probably shaped by a complex interplay between adaptive forces and phylogenetic constraints, the effects of which may vary among different monogenoids (Messu Mandeng *et al.*, 2015; Rodríguez-González *et al.*, 2016, 2017).

Furthermore, research indicates that the intensity and interaction of adaptive forces and phylogenetic constraints can manifest differently in various haptor elements (Vignon *et al.*, 2011; Rodríguez-González *et al.*, 2015). For example, in *Ligophorus cephalis* Rubtsova, Balbuena, Sarabeev, Blasco-Costa & Euzet, 2006 on *Mugil cephalus* (Linnaeus, 1758), a greater control has been observed over the shape and size of the ventral pair of anchors compared to their dorsal counterparts. This difference is noteworthy as the ventral anchors seem responsible for a firmer attachment to the gills (Llopis-Belenguer *et al.*, 2015; Rodríguez-González *et al.*, 2015). In fact, evidence indicates that ventral and dorsal anchors in species of *Ligophorus* and *Cichlidogyrus* exhibit relatively independent evolutionary trajectories, mirroring the functional distinction in their attachment roles (Vignon *et al.*, 2011; Rodríguez-González *et al.*, 2015).

The objectives of the present study were (1) to assess the relative influences of convergence and shared evolutionary history on anchor form on the dactylogyrid parasites of South American

ariids, (2) assess shape and size differences between ventral and dorsal anchors that might provide cues for different functional attachment roles and (3) to use anchor morphology to understand the relationships and evolutionary history of the 3 monogenoid genera studied. Thus, special attention was given to examine whether the morphology of the anchors serves as a basis for synonymizing *Hamatopeduncularia* and *Chauhanellus*, as suggested previously (Kearn and Whittington, 1994; Lim, 1994, 1996). Furthermore, considering the recently suggested ancestral relationship between *Susanlimocotyle* and the latter 2 genera (Soares *et al.*, 2021, 2023b), we also assessed whether there are patterns in the evolutionary changes of anchor morphology within these monogenoids.

## Materials and methods

### Study area, host and parasite samples

The species of fish and parasites from 4 localities in the Brazilian coast (Table 1) are the same used in our previous studies (Soares *et al.*, 2023a, 2023b). The morphological analysis of parasites includes data of all species of *Hamatopeduncularia*, *Chauhanellus* and *Susanlimocotyle* occurring on ariid catfish in the Brazilian coast (10 species in total) (Table 1). A previously published phylogenetic tree based of concatenated partial sequences of genes 18S rDNA, ITS1, 5.8S rDNA and ITS2 performed using Bayesian inference (Soares *et al.*, 2023a) was used to assess the relationships between morphology and evolutionary history of the monogenoid species. The choice of these molecular markers is justified by the relatively large number of sequences available for different species of monogenoid of South American ariid fishes.

### Morphometric data

In landmark-based geometric morphometrics, anatomical landmarks (LMs) are identified and digitized on images of the biological structure under study (Klingenberg, 2010). Herein we placed LMs on the haptor anchors of the monogenoids following Rodríguez-González *et al.* (2017). The haptor of the species studied includes 2 pairs of ventral and dorsal anchors (VA and DA, respectively) (see Fig. 2 in Soares *et al.* (2023a)). We conducted parallel analyses of VA and DA since potential differences between them may provide insights into the distinct selective pressures influencing their morphology due to putative differing roles in attachment (Rodríguez-González *et al.*, 2017). Drawings of VA and DA were taken from the original descriptions of the parasites (holotype) (Domingues and Fehlauer, 2006; Domingues *et al.*, 2016; Soares *et al.*, 2021, 2023b) and were used to place the LMs. One VA and 1 DA of each monogenoid species were processed independently. In each anchor, 5 homologous LMs were placed as per Soares *et al.* (2023a). To capture anchor morphology more accurately, semilandmarks (SLMs) were inserted between each LM (Mitteroecker and Gunz, 2009; Llopis-Belenguer *et al.*, 2015; Rodríguez-González *et al.*, 2015), following the methods for sliding the SLMs (Bookstein *et al.*, 2002). Five groups of 6–29 SLMs were placed equidistantly between LM pairs (for descriptions and locations of LMs and SLMs, see Soares *et al.* (2023a)). The morphology of VA and DA was defined by the Cartesian coordinates ( $x$ ,  $y$ ) of the 83 anatomical points (i.e. LMs and SLMs).

Digitalization of the LMs and SLMs was processed with the TpsDig v2.32 (Rohlf, 2022). Generalized Procrustes analysis in MorphoJ v1.07a (Klingenberg, 2011) was employed to obtain matrices of shape coordinates of VA and DA (datasets 1 and 2, respectively). This analysis removes all information related to position, scale and orientation. Centroid size (CS), estimated as the

**Table 1.** Host species, locality (geographical coordinates) and associated species of *Chauhanellus*, *Hamatopeduncularia* and *Susanlimocotyle* used in the present study

| Host  | Locality   | Parasite   |
|---|--|--|
| <i>Amphiarus rugispinis</i> (Valenciennes, 1840)  | Ajuruteua (0°49'31"N; 46°36'29"W), Bragança, PA, Br          | <i>C. hamatopeduncularoideum</i> Domingues, Soares and Watanabe, 2016, OP681531 <sup>a</sup><br><i>C. neotropicalis</i> Domingues and Fehlauer, 2006   |
| <i>Aspistor luniscutis</i> (Valenciennes, 1840)   | Cananéia (25°02'09.2"S; 47°54'57.8"W), SP, Br                | <i>C. neotropicalis</i> OP681530 <sup>a</sup><br><i>H. cangatae</i> Domingues, Soares and Watanabe, 2016, OP681532 <sup>a</sup>  |
| <i>Aspistor quadriscutis</i> (Valenciennes, 1840) | Ajuruteua (0°49'31"N; 46°36'29"W), Bragança, PA, Br          | <i>C. neotropicalis</i><br><i>H. cangatae</i>  |
| <i>Bagre bagre</i> (Linnaeus, 1766)               | Ajuruteua (0°49'31"N; 46°36'29"W), Bragança, PA, Br          | <i>H. bagre</i> Hargis, 1955, OP681526 <sup>a</sup>  |
| <i>Genidens barbatus</i> (Lacepède, 1803)         | Cananéia (25°02'09.2"S; 47°54'57.8"W), SP, Br                | <i>C. boegeri</i> Domingues and Fehlauer, 2006, OP681529 <sup>a</sup>  |
|   | Estuary of Patos Lagoon (32°08'05.7"S; 52°06'11.2"W), RS, Br | <i>C. boegeri</i><br><i>C. riograndinensis</i> Soares, Martins, Vianna, Domingues and Adriano, 2023b, OP681534 <sup>a</sup>  |
| <i>Genidens genidens</i> (Cuvier, 1829)           | Estuary of Patos Lagoon (32°08'05.7"S; 52°06'11.2"W), RS, Br | <i>C. boegeri</i><br><i>C. riograndinensis</i>   |
| <i>Notarius grandicassis</i> (Valenciennes, 1840) |  | <i>C. neotropicalis</i><br><i>H. cangatae</i>  |
| <i>Sciades couma</i> (Valenciennes, 1840)         | Caratateua (1°59'41.91"S; 46°43'21.385"W), Bragança, PA, Br  | <i>C. hamatopeduncularoideum</i><br><i>C. velum</i> Domingues, Soares and Watanabe, 2016<br><i>C. boegeri</i>  |
| <i>Sciades herzbergii</i> (Bloch, 1794)           | Ajuruteua (0°49'31"N; 46°36'29"W), Bragança, PA, Br          | <i>C. boegeri</i><br><i>C. susamlimae</i> Domingues, Soares and Watanabe, 2016<br><i>C. velum</i> OP681528 <sup>a</sup><br><i>S. narina</i> Soares, Domingues and Adriano, 2021, OP681525 <sup>a</sup> |
| <i>Sciades passany</i> (Valenciennes, 1840)       | Caratateua (1°59'41.91"S; 46°43'21.385"W), Bragança, PA, Br  | <i>C. neotropicalis</i><br><i>C. susamlimae</i> OP681527 <sup>a</sup><br><i>C. velum</i>   |
| <i>Sciades proops</i> (Valenciennes, 1840)        | Ajuruteua (0°49'31"N; 46°36'29"W), Bragança, PA, Br          | <i>C. hypnocleithrum</i> Domingues, Soares and Watanabe, 2016, OP681533 <sup>a</sup>   |

PA, Pará; SP, São Paulo; RS, Rio Grande do Sul; BR, Brazil.

<sup>a</sup>GenBank accession numbers of the DNA sequences of genes 18S rDNA, ITS1, 5.8S rDNA and ITS2 used for the phylogenetic reconstruction of the parasites by Soares *et al.* (2023a). C. = *Chauhanellus*; H. = *Hamatopeduncularia*; S. = *Susanlimocotyle*.

summed squared distances of each LM from the centroid of the form (Zelditch *et al.*, 2012), was used as a measure of anchor size. To visualize interspecific variation in anchor shape, we subjected the matrices of generalized Procrustes analysis coordinates of the VA and DA to principal component analysis (PCA) based on the covariation matrix.

### Assessing the influence of size on anchor shape

We assessed the effects of size on shape variation of the anchors (i.e. interspecific allometry) independently for VA and DA by means of a multivariate regression analysis (Klingenberg, 2016). We used the Procrustes shape coordinates of VA and DA and their log-transformed centroid size (logCS) as input in a multivariate regression through the origin (Lim and Gibson, 2009; Klingenberg *et al.*, 2012). Then, we mapped the residuals from this regression onto the phylogenetic tree of the monogenoids. A sizeable variation between the original datasets and the residuals would suggest that evolutionary allometry (i.e. the allometry between traits measured across species) played an important role in anchor evolution in these monogenoids.

To avoid incorrect interpretations due to a violation of the assumption of independent sampling (Harvey and Pagel, 1991), we also assessed the effect of size on anchor shape with the phylogenetic independent contrast (PIC) correction (Felsenstein, 1985). Since no evidence for allometry to the PIC-corrected analyses was

found ( $P > 0.07$  in VA and DA), the effect of evolutionary allometry was not further considered.

### Evaluating phylogenetic signal in anchor shape and size

To test whether closely related monogenoids tend to have more similar anchors to each other than of more distantly related monogenoids, we evaluated phylogenetic signal in anchor shape and size. For that, we mapped a topology of the phylogenetic tree of our 10 monogenoid species onto the morphospace defined by the 2 first PCA scores (PC1 and PC2) of shapes and onto logCS (anchor size) using squared change-parsimony assuming a model of Brownian-motion (BM) evolution (Klingenberg and Marugán-Lobón, 2013). Phylogenetic signal was evaluated with MorphoJ. Its significance ( $P < 0.05$ ) was established by a permutation test in which the topology was held constant and the principal component scores for each taxon were randomly permuted 10 000 times across the tree (Maddison, 1991; Klingenberg and Gidaszewski, 2010). If there were no correlation between phylogeny and morphometric data, the tree length value should be small (closer to 0 than to 1) and non-significant. Given the disagreement on which approach is more appropriate to measure the phylogenetic signal (Blomberg *et al.*, 2003; Adams, 2014), we also used  $K_{\text{mult}}$  (generalization of Blomberg's  $K$ ) (Adams and Otárola-Castillo, 2013; Adams, 2014) to test for the phylogenetic signal in our data.  $K_{\text{mult}}$  quantifies the extent to which a trait displays phylogenetic signal following BM evolution

(Diniz-Filho *et al.*, 2012).  $K_{\text{mult}} = 0$  suggests no phylogenetic signal,  $K_{\text{mult}} = 1$  indicates that the trait distribution perfectly conforms to BM,  $K_{\text{mult}} < 1$  correspond to trait variation that is larger than expected between taxa of the same lineage and  $K_{\text{mult}} > 1$  indicates stronger similarities among closely related species than expected under BM. The significance of  $K_{\text{mult}}$  ( $P < 0.05$ ) was established based on comparison of the observed value with those obtained in 999 randomizations (Liu *et al.*, 2015). The tests were performed with function *physignal* of the geomorph package v3.0.1 (Adams, 2014) in R v4.1.0 (R Core Team, 2022).

## Results

### Phylogenetic signal in anchor shape and anchor size

The PCA performed with the covariance matrix of LM data of both VA and DA shows that a large part of the variation is contained in relatively few dimensions. The first 2 principal components accounted for 78.5 and 78.3% of the total shape variation in VA and DA, respectively (Table 2). Eigenvalues and variance explained by each principal component are given in Supplementary Table S1.

In the phylomorphospace (Fig. 1a and b), in which the phylogeny of monogenoids (Fig. 1c) was projected onto the morphospace defined by PC1 and PC2 of the VA and DA shape, congeneric species tended to cluster together. Only *Chauhanellus hamatopeduncularoideum* from *Amphiarius rugispinis* and *Sciades couma* did not follow this trend. By contrast, species reported parasitizing the same hosts (i.e. *Chauhanellus boegeri*, *C. velum* and *C. susamlimae* from *Sciades herzbergii*) did not group together in the phylomorphospace. Both approaches, MorphoJ (VA: tree length = 0.249,  $P = 0.007$  and DA: tree length = 0.187,  $P = 0.001$ ), and  $K_{\text{mult}}$  (VA:  $K_{\text{mult}} = 0.78$ ,  $P = 0.01$  and DA:  $K_{\text{mult}} = 1.1$ ,  $P = 0.001$ ) (Fig. 1d–e) supported a significant relationship signal between the shape of VA and DA, and phylogenetic position of the monogenoids.

The deformation grids of each species provide a visual representation of their deviation from the average anchor shape (VA and DA) of the species studied (see parts of anchors in insert in Fig. 1a and b). Based on this evidence, we identify 4 clusters: (i) *Chauhanellus* spp. except *C. hamatopeduncularoideum* (see Table 1 for *Chauhanellus* spp., and Fig. 1c for host–parasite distribution), characterized by a VA with truncated inner root, expanded outer root, short shaft and evenly curved to point; DA with poorly developed inner root, expanded outer root and wide base (Fig. 1a and b, cluster marked with a yellow circle); (ii) *Hamatopeduncularia* spp. (*H. bagre* from *Bagre bagre*; and *H. cangatae* from *A. luniscutis*, *A. quadriscuti* and *N. grandicassis*), characterized by a VA with long inner root, non-expanded outer root, long shaft, curved to point; DA with long inner root, non-differentiated outer root, narrow base (Fig. 1a and b, cluster marked with a green circle); (iii) the monotypic *Susanlimocotyle*

*narina* (from *S. herzbergii*), characterized by a VA with developed inner and outer roots, long shaft, evenly curved to point (*Hamatopeduncularia* morphology); DA with developed inner root, outer root expanded and wide base (*Chauhanellus* morphology) (Fig. 1a and b, cluster marked with a red circle), with characteristics intermediate between cluster ii (at the VA) and between clusters i and ii (at the DA), which seem to represent the characteristics shared with the species of *Hamatopeduncularia* and *Chauhanellus*, consistent with the close relationship of these species suggested by the phylogenetic tree (Fig. 1c); and (iv) *C. hamatopeduncularoideum* (from *A. rugispinis* and *S. couma*), characterized by a VA with long inner root, not expanded outer root, long shaft, curved to point (*Hamatopeduncularia* morphology); DA with poorly developed inner root, expanded outer root and wide base (*Chauhanellus* morphology), with intermediate characteristics between cluster ii (at the VA) and between i (at the DA), and *C. hamatopeduncularoideum* covering the same swath in phylomorphospace for VA and DA (see Fig. 1a and b, cluster marked with a pink circle).

The phylogeny of monogenoids projected onto the morphospace defined by allometry-free (size-corrected) PC1 and PC2 of anchor shape yielded a tree length of 0.02 for VA and DA (Fig. 2a and b). The multivariate regression of Procrustes coordinates on logCS, provided evidence for an allometric relationship between shape and size only for DA (VA:  $P = 0.5$ ; DA:  $P = 0.01$ ), accounting for 29.7% of the total shape variation of DA. Phylogenetic signal was again highly significant, both in MorphoJ ( $P = 0.007$  each) and  $K_{\text{mult}}$  (size-corrected) (VA:  $K_{\text{mult}} = 0.72$ ,  $P = 0.009$  and DA:  $K_{\text{mult}} = 0.98$ ,  $P = 0.001$ ). The scatter graph of VA (Fig. 2a) showed small branches of *C. susamlimae*, *C. velum*, *C. riograndinensis* and *H. bagre* than in the PCA uncorrected for size (Fig. 1a). By contrast, the branches of *C. boegeri*, *C. neotropicalis*, *C. susamlimae*, *C. velum* and *C. riograndinensis* were larger than the original PCA in the DA scatterplot (compare Fig. 1b with Fig. 2b). Whereas the position of species in the original and size-corrected was similar in the VA phylospaces, it was not the case in the DA phylospaces. Consequently, allometry had a significant effect on the overall variation of DA shape, but not on VA shape.

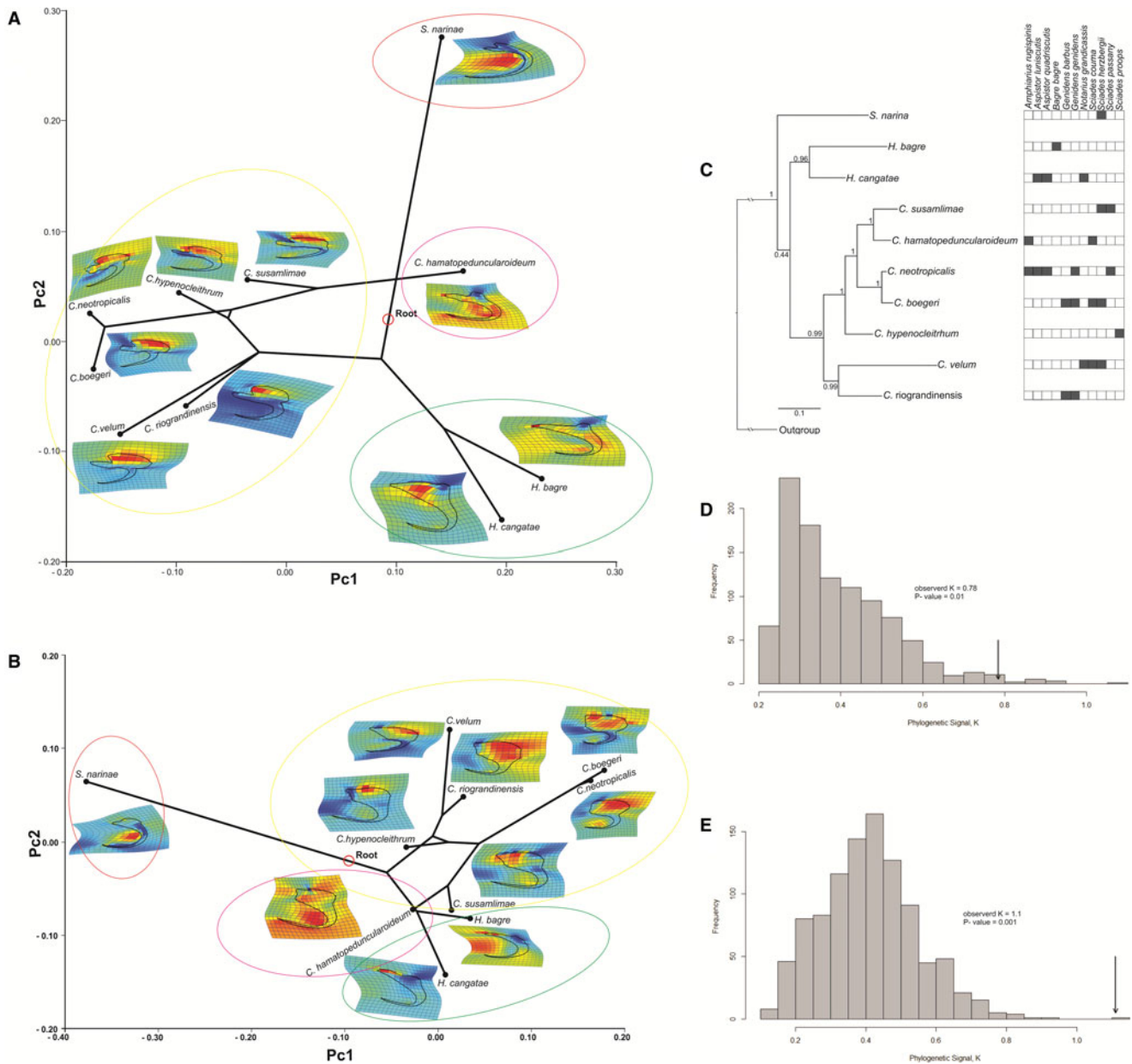
In Fig. 3, the molecular phylogeny projected onto the gradient in size (logCS) of VA and DA is shown along with the cumulative branch length from the root of the tree. This mapping resulted in tree lengths of VA and DA of 0.34 and 0.62, respectively, computed in units of logCS distance along all branches. Phylogenetic signal tested in MorphoJ by random permutation of logCS was not statistically significant in both anchors (VA:  $P = 0.07$  and DA:  $P = 0.12$ ). However,  $K_{\text{mult}}$  indicated a significant phylogenetic signal in VA but not in DA (VA:  $K_{\text{mult}} = 0.99$ ,  $P = 0.03$ ; DA:  $K_{\text{mult}} = 0.68$ ,  $P = 0.11$ ).

## Discussion

Monogenoids of the genera *Hamatopeduncularia*, *Chauhanellus* and *Susanlimocotyle*, parasitic on South American ariids, exhibit distinct variations in anchor shape (Domingues *et al.*, 2016; Soares *et al.*, 2021, 2023b), and these differences are clearly reflected in their positions within the phylomorphospace (Fig. 1a and b). Thus, it is not surprising that for the 3 genetic lineages (*Susanlimocotyle*, *Hamatopeduncularia* and *Chauhanellus*) (Fig. 1c), the VA and DA shape exhibit a significant phylogenetic signal, suggesting that evolutionary history played an important role in determining the shape of haptor anchors. This agrees with other studies, which suggest a consistent relationship between anchor morphology and phylogeny in monogenoids (Sarabeev and Desdevises, 2014; Khang *et al.*, 2016; Rodríguez-González *et al.*, 2017).

**Table 2.** PCA of variation among the shapes of species for ventral and dorsal anchors of monogenoids from ariids

| Anchor  | Eigenvalue            | Total variance (%) |
|---------|-----------------------|--------------------|
| Ventral |                       |                    |
| PC1     | $2.68 \times 10^{-2}$ | 49.8               |
| PC2     | $1.54 \times 10^{-2}$ | 28.7               |
| Dorsal  |                       |                    |
| PC1     | $2.27 \times 10^{-2}$ | 58.6               |
| PC2     | $7.64 \times 10^{-3}$ | 19.7               |



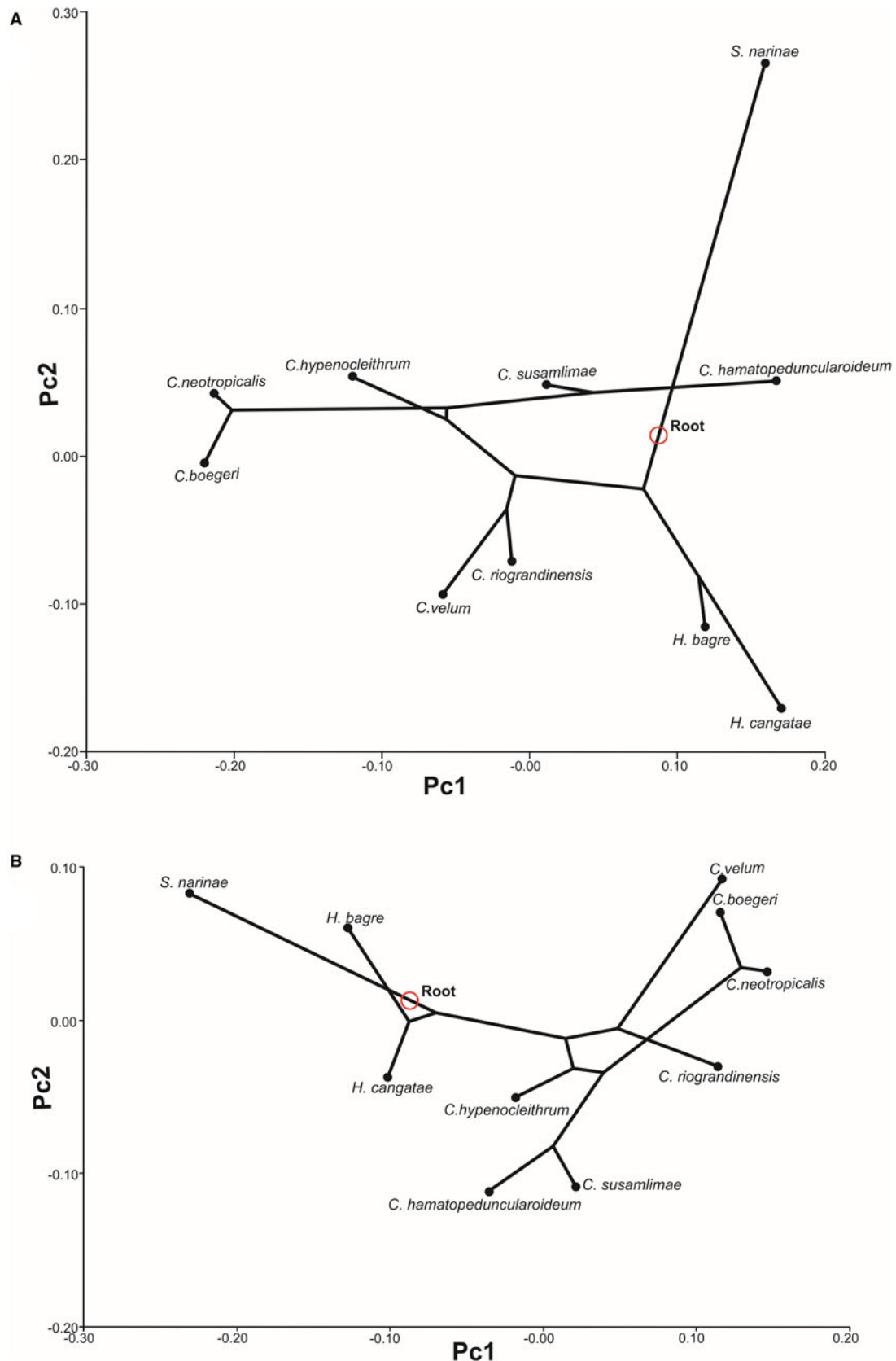
**Figure 1.** Phylomorphospace showing changes in the shape of ventral (a) and (b) dorsal haptor anchors of the species of monogenoids (*Susanlimocotyle*, *Hamatopeduncularia* and *Chauhanellus* spp.) studied mapped onto phylogeny. Estimated changes in anchor shapes are shown as thin-plate-spline deformation grids with colour-scaled coded Jacobian expansion factors (red for factors  $>1$ , indicating expansion; strong blue for factors between 0 and 1, indicating contraction) were used. The insert shows the parts of an anchor in monogenoids species from ariids. *S.*, *Susanlimocotyle*; *H.*, *Hamatopeduncularia*; *C.*, *Chauhanellus*; The coloured circles represent the clusters formed. (c) Bayesian tree based on partial sequences of genes 18S rDNA, ITS1, 5.8S rDNA and ITS2 sequences of representative individuals of 10 monogenoids species of the genera *Susanlimocotyle*, *Hamatopeduncularia* and *Chauhanellus* (posterior probabilities support values are given above the branches); and host-parasite distribution. (d–e) Histogram of  $K_{mult}$  values obtained from 999 permutations of the ventral (d) and (e) dorsal anchors shape data and the phylogeny, with the position of observed value of  $K_{mult}$  identified.

Visually, the distribution of anchors in phylomorphospace was more convincing for the VA than for DA, indicating the formation of clusters (Fig. 1a and b), especially regarding the separation of *Hamatopeduncularia* and *Chauhanellus* lineages into 2 distinct clusters. Interestingly, the  $K_{mult}$  corresponding to shape was  $<1$  for VA and  $>1$  for DA (Fig. 1d–e). Thus, the phenotypic variation in VA is greater than expected between taxa of the same lineage (Adams, 2014). This indicates that the evolutionary processes acting on VA shape did not act with similar intensity, or were not the same as in DA in these genera.

In addition, the deformation grids plotted in Fig. 1a indicate that the inner and outer roots of the VA are more differentiated than those of the DA counterparts in species of each genus (Fig. 1b). In general, the anchor roots of Dactylogyridae are the

point of connection and articulation with the bars, aided by the insertions of the haptor muscles. Functionally, they have the role of controlling the intensity of attachment to the host (Kearn, 1994). Thus, the differences observed between VA and DA can be explained in terms of different functional roles of these structures, which seems to be common in Dactylogyridae (Vignon *et al.*, 2011; Llopis-Belenguer *et al.*, 2015; Rodríguez-González *et al.*, 2015).

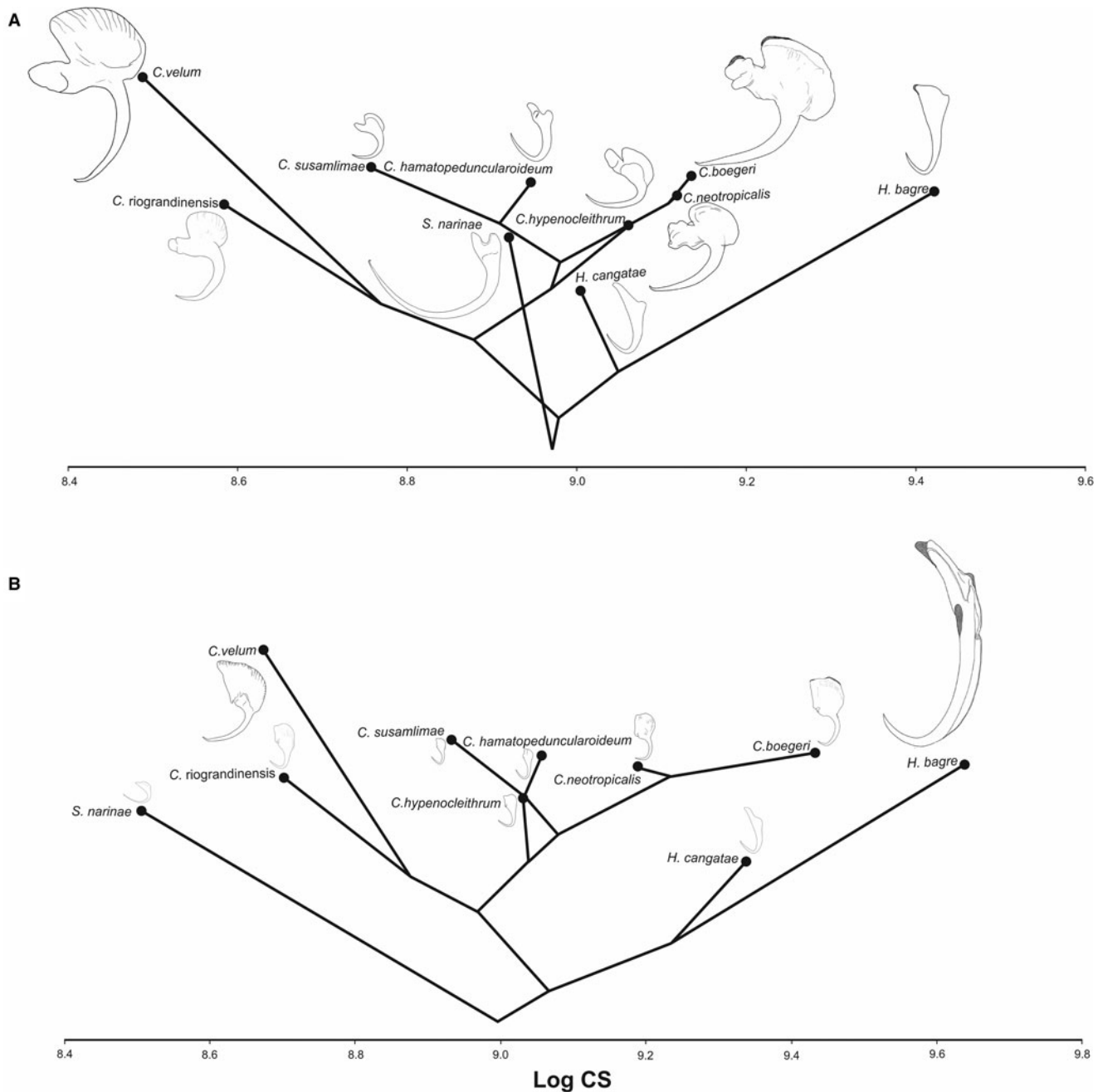
Interestingly, the VA shape in *C. hamatopeduncularoideum*, characterized by an inner root long, outer root not expanded, long shaft, curved to point, exhibits a greater resemblance in the phylomorphospace to those found in *Hamatopeduncularia* species rather than other *Chauhanellus* species (Fig. 1a). One could speculate that similarity in the shape of the anchors may result from



**Figure 2.** Phylomorphospace showing changes in shape (corrected for size) of ventral (a) and (b) dorsal haptor anchors of the species of monogenoids (*Susanlimocotyle*, *Hamatopeduncularia* and *Chauhanellus* spp.) studied mapped onto phylogeny. Species abbreviations as in Fig. 1.

adaptation to the gill morphology of the host. However, *C. hamatopeduncularoideum* does not share any host with *Hamatopeduncularia* spp. (Fig. 1c). Alternatively, *C. hamatopeduncularoideum* could be considered a species of *Hamatopeduncularia*. In fact, some authors (Kearn and Whittington, 1994; Lim, 1994, 1996;

Lim *et al.*, 2001; Domingues *et al.*, 2016) have suggested that *Chauhanellus* and *Hamatopeduncularia* may be considered synonyms, because certain species within each genus share morphological characteristics that were originally used to differentiate the 2 genera in the past (Soares *et al.*, 2021, 2023b). However, evidence



**Figure 3.** Projection of phylogenetic tree from monogenoids (*Susanlimocotyle*, *Hamatopeduncularia* and *Chauhanellus* spp.) onto log centroid size (logCS) of ventral (a) and dorsal (b) haptor anchors. Species abbreviations as in Fig. 1. The anchors displayed are scaled as per the logCS scale to convey the gradient in size.

from molecular data does not support neither the inclusion of *C. hamatopeduncularoideum* within *Hamatopeduncularia*, nor the synonymy of the 2 genera (Fig. 1c) (Soares *et al.*, 2021, 2023b).

Alternatively, the presence of *Hamatopeduncularia*-like anchors in *Chauhanellus* species suggests that this morphology might have been present in the common ancestor of both genera (Kearn and Whittington, 1994). Thus, reversal to the ancestral character state would account for the anchor morphology in *C. hamatopeduncularoideum*, as suggested for other structures in some monogenoids (Šimková *et al.*, 2006).

Šimková *et al.* (2006) carried out ancestral state reconstructions on *Dactylogyrus* species, revealing a shift in haptor anchor shapes from type 2 to type 1 for the majority of species (see Šimková *et al.*, 2006, p. 1028, Table 1 for anchor shapes). However, the authors noted a reversion to the ancestral state in derivative

species. This observation appears to be applicable to the *Hamatopeduncularia*-type anchors in *C. hamatopeduncularoideum*.

Indeed, the VA shape of *S. narinae* also conforms to that of *Hamatopeduncularia* (see parts of anchors in insert in Fig. 1a), whereas the DA shape seems a composite of those of *Hamatopeduncularia* and *Chauhanellus* (see parts of anchors in insert in Fig. 1b). This suggests that some character states of *Susanlimocotyle* shared *Hamatopeduncularia* and *Chauhanellus* might represent evolutionary ancestral forms (Fig. 1a–c).

An additional feature of *C. hamatopeduncularoideum* shared with *Hamatopeduncularia* spp. is the presence of a digitiform haptor. This character was originally considered as diagnostic of *Hamatopeduncularia* (Yamaguti, 1953), but it was later found in some *Chauhanellus* spp. (e.g. *C. susamlimae* and *C. riograndinensis*) (Kearn and Whittington, 1994; Lim, 1994, 1996; Lim *et al.*, 2001; Domingues *et al.*, 2016; Soares *et al.*, 2023b).

Interestingly, the *Chauhanellus* species that have a digitiform haptor (*C. hamatopeduncularoideum*, *C. susamlimae* and *C. riograndinensis*) have smaller anchors than closely related congeneric species (Fig. 3). This could account for the lack of significant phylogenetic signal in anchor size found in most analyses. Only the  $K_{\text{mult}}$  result for DA was significant, but Fig. 3b suggests that the relationship between size and phylogenetic relatedness is not strong.

Kearn and Whittington (1994) suggested that an important innovation in some ancestral ancyrocephalines (Dactylogyridae) was the development of hooklet-bearing papillae (=digitiform haptor) with the ability to elongate. According to these authors, this provided these parasites with multiple attachment points to the gills, and offered little resistance, minimizing the threats of displacement by water currents, allowing more versatility in attachment sites. Thus, the digitiform haptor decreased the function of the anchors, which resulted in a reduction of their size (Kearn and Whittington, 1994). This scenario is supported by the present study, as the presence of a digitiform haptor in *C. hamatopeduncularoideum*, *C. susamlimae* and *C. riograndinensis* coincides with a reduction in anchor size. Likewise, the digitiform haptor may have caused the secondary loss of anchors in other dactylogirid genera (e.g. *Trinigyryus* Hanek, Molnar & Fernando (1974) sensu Kritsky *et al.* (1986)). However, in other genera with no digitiform haptor, like *Dactylogyryus* and *Dogielius*, the ventral bar is significantly reduced and ventral anchors are altogether absent (Pravdová *et al.*, 2018). Thus, other selective forces would account for the reduction of haptors elements. In any case, this observation strengthens the proposition that dorsal and ventral anchors may undergo distinct evolutionary trajectories within the Dactylogyridae.

## Conclusion

Our study suggests that phylogeny has driven the evolution of shape but not size of the anchors of monogenoids from South American ariids. However, it seems that the emergence of the digitiform haptor in *Hamatopeduncularia* and some species of *Chauhanellus* played an important role in the reduction of anchors, as suggested by other authors, and may account for secondary losses in other groups of monogenoids.

Nevertheless, we acknowledge the limited scope of our study. While typical geometric morphometric studies analyse several specimens per species, our investigation employed a single representative per species due to lack of specimen availability. Our approach rest on the assumption that intraspecific variation and measurement error are smaller than interspecific differences (Klingenberg and Marugán-Lobón, 2013). This premise appears substantiated given that morphological differences in both size and shape of the anchors between species were clear and substantial (Fig. 3). Nevertheless, we must acknowledge the potential impact of even minor levels of intraspecific variation or measurement error on the conclusions drawn from our study. Hence, while our findings offer valuable insights into the phylogenetic effects on anchor form, future research incorporating multiple specimens will undoubtedly contribute to a more nuanced picture of the evolution of anchor morphology of monogenoids of ariid fishes.

In addition, future studies should also use molecular markers from different regions (i.e. 28S rDNA, COI) and include a wider range of taxa, including the type species of each genus (*Hamatopeduncularia arii* Yamaguti, 1953 and *Chauhanellus oculatus* Bychowsky & Nagibina, 1969) and representatives of New-World and Old-World lineages as proposed by Soares *et al.* (2023b).

**Supplementary material.** The supplementary material for this article can be found at <https://doi.org/10.1017/S0031182024000192>

**Data availability statement.** Datasets 1–2 required to perform all the analyses are deposited on Zenodo (<https://zenodo.org/records/10412631>).

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## References

- Adams DC (2014) A generalized K statistic for estimating phylogenetic signal from shape and other high-dimensional multivariate data. *Systematic Biology* **63**, 685–697.
- Adams DC and Otarola-Castillo E (2013) Geomorph: an R package for the collection and analysis of geometric morphometric shape data. *Methods Ecology and Evolution* **4**, 393–399.
- Adams DC, Rohlf FJ and Slice DE (2004) Geometric morphometrics: ten years of progress following the ‘revolution’. *Italian Journal of Zoology* **71**, 5–16.
- Adams DC, Rohlf FJ and Slice DE (2013) A field comes of age: geometric morphometrics in the 21st century. *Hystrix* **24**, 7–14.
- Baillie C, Welicky RL, Hadfield KA, Smit NJ, Mariani S and Beck RMD (2019) Hooked on you: shape of attachment structures in cymothoid isopods reflects parasitic strategy. *BMC Evolutionary Biology* **19**, 207.
- Blomberg SP, Garland T and Ives AR (2003) Testing for phylogenetic signal in comparative data: behavioral traits are more labile. *Evolution* **57**, 717–745.
- Boeger WA and Vianna RT (2006) Monogenoidea. In Thatcher VE (ed.), *Aquatic Biodiversity in Latin America: Amazon Fish Parasites*. Sofia, Bulgaria: Pensoft Publishers, pp. 42–116.
- Bookstein FL, Streissguth AP, Sampson PD, Connor PD and Barr HM (2002) Corpus callosum shape and neuropsychological deficits in adult males with heavy fetal alcohol exposure. *Neuroimage* **15**, 233–251.
- Bychowsky BE (1957) *Monogenetic Trematodes. Their Systematic and Phylogeny*. Moscow, USSR: Academy of Sciences of the USSR.
- Cruz-Laufer AJ, Pariselle A, Jorissen MWP, Muterezi Bukinga F, Al Assadi A, Van Steenberge M, Koblmüller S, Sturmbauer C, Smeets K, Huysse T, Artois T and Vanhove MPM (2022) Somewhere I belong: phylogeny and morphological evolution in a species-rich lineage of ectoparasitic flatworms infecting cichlid fishes. *Cladistics* **38**, 465–512.
- Desdevises Y, Morand S, Jousson O and Legendre P (2002) Coevolution between *Lamellodiscus* (Monogenea; Diplectanidae) and Sparidae (Teleostei): the study of a complex host-parasite system. *Evolution* **56**, 2459–2471.
- Diniz-Filho JA, Rangel TF, Santos T and Bini LM (2012) Exploring patterns of interspecific variation in quantitative traits using sequential phylogenetic eigenvector regressions. *Evolution* **66**, 1079–1090.
- Domingues MV and Fehlaue KH (2006) New species of *Chauhanellus* (Monogenoidea, Platyhelminthes) from the gills of Southern Atlantic marine catfishes (Siluriformes, Ariidae) on the Neotropical Region. *Zootaxa* **1365**, 61–68.
- Domingues MV, Soares GB and Watanabe A (2016) Monogenoidea (Polyonchoinea: Dactylogyridae) parasitizing the gills of marine catfish



- (Siluriformes: Ariidae) inhabiting the Atlantic Amazon Coast of Brazil. *Zootaxa* **4127**, 301–326.
- Felsenstein J** (1985) Phylogenies and the comparative method. *American Naturalist* **125**, 1–15.
- Friedman ST, Martinez CM, Price AS and Wainwright PC** (2019) The influence of size on body shape diversification across Indo-Pacific shore fishes. *Evolution* **73**, 1873–1884.
- Harvey PH and Pagel MD** (1991) *The Comparative Method in Evolutionary Biology*. Oxford, UK: Oxford University Press.
- Kearn GC** (1994) Evolutionary expansion of the Monogenea. *International Journal for Parasitology* **24**, 1227–1271.
- Kearn GC and Whittington ID** (1994) Ancyrocephaline monogeneans of the genera *Chauhanellus* and *Hamatopeduncularia* from the gills of the blue catfish, *Arius graeffei*, in the Brisbane river and Moreton bay, Queensland, Australia, with descriptions of four new species. *International Journal for Parasitology* **24**, 569–588.
- Kerschbaumer M and Pfingstl T** (2021) Testing for phylogenetic signal in claws suggests great influence of ecology on Caribbean intertidal arthropods (Acari, Oribatida). *Scientific Reports* **11**, 4398.
- Khang TF, Soo OYM, Tan WB and Lim LHS** (2016) Monogenean anchor morphology: systematic value, phylogenetic signal, and evolution. *PeerJ* **4**, e1668.
- Klingenberg CP** (2010) Evolution and development of shape: integrating quantitative approaches. *Nature Reviews Genetics* **11**, 623–635.
- Klingenberg CP** (2011) MorphoJ: an integrated software package for geometric morphometrics. *Molecular Ecology Resources* **11**, 353–357.
- Klingenberg CP** (2016) Size, shape, and form: concepts of allometry in geometric morphometrics. *Development Genes and Evolution* **226**, 113–137.
- Klingenberg CP and Gidaszewski NA** (2010) Testing and quantifying phylogenetic signals and homoplasy in morphometric data. *Systematic Biology* **59**, 245–261.
- Klingenberg CP and Marugán-Lobón J** (2013) Evolutionary covariation in geometric morphometric data: analyzing integration, modularity, and allometry in a phylogenetic context. *Systematic Biology* **62**, 591–610.
- Klingenberg CP, Duttke S, Whelan S and Kim M** (2012) Developmental plasticity, morphological variation and evolvability: a multilevel analysis of morphometric integration in the shape of compound leaves. *Journal of Evolutionary Biology* **25**, 115–129.
- Kmentová N, Koblmüller S, Van Steenberghe M, Artois T, Muterezi Bukinga F, Mulimbwa N'sibula T, Muzumani Risasi D, Masilya Mulungula P, Gelnar M and Vanhove MPM** (2020) Failure to diverge in African Great Lakes: the case of *Doliceiroplectanum lacustre* gen. nov. comb. nov. (Monogenea, Diplectanidae) infecting latid hosts. *Journal of Great Lakes Research* **46**, 1113–1130.
- Kritsky DC, Boeger WA and Thatcher VE** (1986) Neotropical Monogenea. 9. Status of *Trinigyrus* Hanek, Molnar & Fernando, 1974 (Dactylogyridae) with descriptions of two new species from loriciid catfishes from the Brazilian Amazon. *Proceedings of the Biological Society of Washington* **99**, 392–398.
- Lim LHS** (1994) *Chauhanellus* Bychowsky & Nagibina, 1969 (Monogenea) from ariid fishes (Siluriformes) of peninsular Malaysia. *Systematic Parasitology* **28**, 99–124.
- Lim LHS** (1996) Eight new species of *Hamatopeduncularia* Yamaguti, 1953 (Monogenea: Ancyrocephalidae) from Ariidae of peninsular Malaysia. *Systematic Parasitology* **33**, 53–71.
- Lim LHS and Gibson DI** (2009) A new monogenean genus from an ehippid fish off Peninsular Malaysia. *Systematic Parasitology* **73**, 13–25.
- Lim LHS, Timofeeva TA and Gibson DI** (2001) Dactylogyridean monogeneans of the siluriform fishes of the old world. *Systematic Parasitology* **50**, 159–197.
- Liu H, Xu Q, He P, Santiago LS, Yang K and Ye Q** (2015) Strong phylogenetic signals and phylogenetic niche conservatism in ecophysiological traits across divergent lineages of Magnoliaceae. *Scientific Reports* **5**, 12246.
- Llopis-Belenguer C, Balbuena JA, Galván-Femenía I and Rodríguez-González A** (2015) Phenotypic buffering in a monogenean: canalization and developmental stability in shape and size of the haptor anchors of *Ligophorus cephalis* (Monogenea: Dactylogyridae). *PLoS ONE* **10**, e0142365.
- Maddison WP** (1991) Squared-change parsimony reconstructions of ancestral states for continuous-valued characters on a phylogenetic tree. *Systematic Zoology* **40**, 304–314.
- Mendlová M and Šimková A** (2014) Evolution of host specificity in monogeneans parasitizing African cichlid fish. *Parasites and Vectors* **7**, 69.
- Messu Mandeng FD, Bilong Bilong CF, Pariselle A, Vanhove MPM, Bija Nyom AR and Agnèse JF** (2015) A phylogeny of *Cichlidogyrus* spp. (Monogenea, Dactylogyridae) clarifies a host-switch between fish families and reveals an adaptive component to attachment organ morphology of this parasite genus. *Parasites & Vectors* **8**, 582.
- Míguez-Lozano R, Rodríguez-González A and Balbuena JA** (2017) A quantitative evaluation of host-parasite coevolutionary events in three genera of monopisthocotylean monogeneans. *Vie et Milieu – Life and Environment* **67**, 103–119.
- Mitteroecker P and Gunz P** (2009) Advances in geometric morphometrics. *Evolutionary Biology* **36**, 235–247.
- Mladineo I, Šegvić-Bubić T, Stanić R and Desdevises Y** (2013) Morphological plasticity and phylogeny in a monogenean parasite transferring between wild and reared fish populations. *PLoS ONE* **8**, e62011.
- Poisot T, Verneau O and Desdevises Y** (2011) Morphological and molecular evolution are not linked in *Lamellogadus* (Platyhelminthes, Monogenea). *PLoS ONE* **6**, e26252.
- Poulin R** (2002) The evolution of monogenean diversity. *International Journal for Parasitology* **32**, 245–254.
- Pravdová M, Ondračková M, Přikrylová I, Blažek R, Mahmoud Z and Gelnar M** (2018) Dactylogyrids (Platyhelminthes: Monogenea) from Sudanese *Labeo* spp., with a description of *Dogielius sennarensis* n. sp. and a redescription of *Dogielius flosculus* Guégan, Lambert & Euzet, 1989. *Helminthologia* **55**, 306–321.
- R Core Team** (2022) *R: A Language and Environment for Statistical Computing*. Vienna, Austria: R Foundation for Statistical Computing. Available at <https://www.r-project.org/> (Accessed 03 march 2022).
- Rahmouni C, Van Steenberghe M, Vanhove MPM and Šimková A** (2021) Intraspecific morphological variation in *Cichlidogyrus* (Monogenea) parasitizing two cichlid hosts from Lake Tanganyika exhibiting different dispersal capacities. *Hydrobiologia* **848**, 3833–3845.
- Rahmouni C, Vanhove MPM, Koblmüller S and Šimková A** (2022) Molecular phylogeny and speciation patterns in host-specific monogeneans (*Cichlidogyrus*, Dactylogyridae) parasitizing cichlid fishes (Cichliformes, Cichlidae) in Lake Tanganyika. *International Journal for Parasitology* **52**, 359–375.
- Rodríguez-González A, Míguez-Lozano R, Llopis-Belenguer C and Balbuena JA** (2015) Phenotypic plasticity in haptor structures of *Ligophorus cephalis* (Monogenea: Dactylogyridae) on the flathead mullet (*Mugil cephalus*): a geometric morphometric approach. *International Journal for Parasitology* **45**, 295–303.
- Rodríguez-González A, Míguez-Lozano R, Sarabeev V and Balbuena JA** (2016) Evolutionary modularity and morphological integration in the haptor anchor structures of *Ligophorus* spp. (Monogenea: Dactylogyridae). *Parasitology Research* **115**, 3519–3533.
- Rodríguez-González A, Sarabeev V and Balbuena JA** (2017) Evolutionary morphology in shape and size of haptor anchors in 14 *Ligophorus* spp. (Monogenea: Dactylogyridae). Evolutionary morphology in shape and size of haptor anchors in 14 *Ligophorus* spp. (Monogenea: Dactylogyridae). *PLoS ONE* **12**, e0178367.
- Rohlf FJ** (2022). Tps Series. Department of Ecology and Evolution, State University of New York, Stony Brook, New York. Available at <http://sbmorphometrics.org/index.html> (Accessed 03 march 2022).
- Sarabeev V and Desdevises Y** (2014) Phylogeny of the Atlantic and Pacific species of *Ligophorus* (Monogenea: Dactylogyridae): morphology vs molecules. *Parasitology International* **63**, 9–20.
- Seidlová L, Benovics M and Šimková A** (2022) Gill monogeneans of Neotropical cichlid fish: diversity, phylogenetic relationships, and host-parasite cophylogenetic associations. *International Journal for Parasitology* **52**, 603–615.
- Šimková A and Morand S** (2008) Co-evolutionary patterns in congeneric monogeneans: a review of *Dactylogyrus* species and their cyprinid hosts. *Journal of Fish Biology* **73**, 2210–2227.
- Šimková A, Ondračková M, Gelnar M and Morand S** (2002) Morphology and coexistence of congeneric ectoparasite species: reinforcement of reproductive isolation? *Biological Journal of the Linnean Society* **76**, 125–135.
- Šimková A, Verneau O, Gelnar M and Morand S** (2006) Specificity and specialization of congeneric monogeneans parasitizing cyprinid fish. *Evolution* **60**, 1023–1037.
- Soares GB, Domingues MV and Adriano EA** (2021) An integrative taxonomic study of *Susanlimocotyle narina* n. gen. n. sp. (Monogenea, Dactylogyridae) from the nasal cavities of a marine catfish (Siluriformes, Ariidae) from the Atlantic Amazon Coast of Brazil and new molecular data of *Chauhanellus* spp. *Parasitology International* **81**, 102271.

- Soares GB, Adriano EA, Domingues MV and Balbuena JA** (2023a) Diversification processes between monogenoids (Dactylogyridae) and their marine catfish (Siluriformes: Ariidae) from the Atlantic coast of South America. *Parasitology* **150**, 184–194.
- Soares GB, Martins MF, Vianna RT, Domingues MV and Adriano EA** (2023b) Morphology and molecular phylogeny of *Chauhanellus* Bychowsky & Nagibina, 1969 (Monogenoidea) parasitizing marine catfish (Ariidae) from the Atlantic coast of South America: a new species, supplementary taxonomic information, and new insights. *Journal of Helminthology* **97**, 1–14.
- Vanhove MPM and Huysse T** (2015) Host specificity and species jumps in fish-parasite systems. In Morand S, Krasnov BR and Littlewood DTJ (eds), *Parasite Diversity and Diversification*. Cambridge, UK: Cambridge University Press, pp. 401–419.
- Vanhove MPM, Pariselle A, Van Steenberge M, Raeymaekers JAM, Hablützel PI, Gillardin C, Hellemans B, Breman FC, Koblmüller S, Sturmbauer C, Snoeks J, Volckaert FAM and Huysse T** (2015) Hidden biodiversity in an ancient lake: phylogenetic congruence between Lake Tanganyika tropheine cichlids and their monogenean flatworm parasites. *Scientific Reports* **5**, 13669.
- Vignon M** (2011) Inference in morphological taxonomy using collinear data and small sample sizes: monogenean sclerites (Platyhelminthes) as a case study. *Zoologica Scripta* **40**, 306–316.
- Vignon M, Pariselle A and Vanhove MPM** (2011) Modularity in attachment organs of African *Cichlidogyrus* (Platyhelminthes: Monogenea: Ancyrocephalidae) reflects phylogeny rather than host specificity or geographic distribution. *Biological Journal of the Linnean Society* **102**, 694–706.
- Whittington ID** (2005) Monogenea monopisthocotylea (ectoparasitic flukes). In Rohde K (ed.), *Marine Parasitology*. Victoria, Australia: CSIRO Publishing, pp. 63–72.
- Whittington ID, Cribb BW, Hamwood TE and Halliday JA** (2000) Host-specificity of monogenean (platyhelminth) parasites: a role for anterior adhesive areas? *International Journal for Parasitology* **30**, 305–320.
- Yamaguti S** (1953) Parasitic worms mainly from Celebes. Part 2. Monogenetic trematodes of fishes. *Acta Medica Okawama* **8**, 203–256.
- Zelditch ML, Swiderski DL and Sheets HD** (2012) *Geometric Morphometrics for Biologists: A Primer*, 2nd Edn. Amsterdam, Netherlands: Academic Press.